

Available online at www.sciencedirect.com



Journal of Nutritional Biochemistry

Journal of Nutritional Biochemistry 21 (2010) 255-260

REVIEWS: CURRENT TOPICS

Mechanisms of anti-atherosclerotic functions of soy-based diets $\stackrel{\leftrightarrow}{\sim}$

Shanmugam Nagarajan*

Department of Microbiology and Immunology, Arkansas Children's Nutrition Center, University of Arkansas for Medical Sciences, Little Rock, AR 72202, USA

Received 12 February 2009; received in revised form 17 July 2009; accepted 3 September 2009

Abstract

Soy-based diets have been reported to protect against the development of atherosclerosis. However, the underlying mechanism(s) for this protection remains unknown. Although atherosclerosis was traditionally considered a disease associated with impaired lipid metabolism, in recent years the inflammatory components of atherosclerosis have been explored. Recent studies have convincingly delineated that uncontrolled chronic inflammation is the principal contributing factor for the initiation and progression of atherosclerosis. Interaction between activated monocytes and vascular endothelial cells is an early event in atherogenesis. The adhesion of leukocytes, including monocytes, to the inflamed-vascular endothelium and their transmigration into intima initiate the inflammatory processes. Following transmigration, monocytes in the intima are transformed to macrophages, which take up oxidized-LDL (oxLDL) to generate lipid-laden macrophages, also known as foam cells. Hence, in this review article the inflammatory processes associated with atherosclerosis and possible anti-inflammatory functions of soy-based diets contributing to the prevention of atherosclerosis are presented. © 2010 Elsevier Inc. All rights reserved.

Keywords: Soy; Atherosclerosis; Isoflavones; Inflammation; Cell adhesion

1. Introduction

Cardiovascular disease (CVD) continues to be the leading cause of death in the US and other Western countries. Atherosclerosis, thickening of arterial intima, is one of the most common causes of CVD. Recent studies have suggested that maternal hypercholesterolemia during pregnancy is associated with a marked increase in aortic fatty streak formation in human fetuses and a more rapid progression of atherosclerosis during normocholesterolemic childhood [1–4]. Furthermore, with the increasing incidence of type II diabetes and childhood obesity, atherosclerosis has become a major health problem in the US and in many developing nations [5,6]. Hence, the prevention of atherosclerosis by dietary intervention would reduce the early onset and incidence of CVD.

* Tel.: +1 501 364 2814; fax: +1 501 364 3161.

E-mail address: nagarajanshanmugam@uams.edu.

1.1. Soy and cardiovascular disease

Epidemiological studies have shown a lower incidence of CVD in Asia than in Western countries [7,8] that is associated with a reduced mortality rate due to coronary heart disease in populations consuming soy [9]. These studies have suggested that consumption of a traditional Asian diet high in soy may play a pivotal role in prevention of chronic diseases such as atherosclerosis [7]. The atheroprotective effects of soy-based diets have been attributed to its effect on reducing serum cholesterol levels in human nutrition studies [10–14]. Similar findings have also been reported in nonhuman primates fed soy-based diets compared to those fed control diets [15-17]. However, recent studies in the atherosclerosis-susceptible apolipoprotein E knockout (apoE-/-) mouse models [18,19] showed that atherosclerotic lesions are reduced when fed a soy-containing diet despite unchanged serum lipid levels [20-22]. These studies suggest that dietary soy may inhibit atherosclerotic lesion development by mechanism(s) other than lowering serum cholesterol levels. However, the mechanism(s) and component(s) of soy responsible for these effects remain to be resolved.

1.2. Atherosclerosis is a chronic inflammatory disease

Recent studies have unequivocally demonstrated that atherosclerosis is a chronic inflammatory disease [23,24], and vascular inflammation has been identified as an early event in the pathogenesis of atherosclerosis [23,24]. Four early cellular events contribute to

Abbreviations: ApoE—/—, apolipoprotein E knockout; CD54, intercellular adhesion molecule-1; CD62E, E-selectin; CD62P, P-selectin; CD106, vascular cell adhesion molecule-1; CVD, cardiovascular disease; LDL, low-density lipoprotein; MCP-1, monocyte chemoattractant protein-1; oxLDL, oxidized-LDL; PPAR, peroxisome proliferator-activated receptor; TF, tissue factor.

 $[\]stackrel{\text{\tiny{th}}}{\longrightarrow}$ This work was supported by a grant from USDA (CRIS 6251-51000-005-00D) (SN).

the initiation of inflammatory processes associated with atherosclerosis: (1) endothelial cell activation resulting in enhanced adhesive property of vascular endothelial cells, (2) activation of monocytes leading to the firm adhesion of circulating inflammatory cells, (3) transmigration of monocytes to the intima where these cells transform to become macrophages, and, finally, (4) uptake of modified low-density lipoprotein (LDL), such as an oxidized form of LDL (oxLDL), by macrophages resulting in transformation of macrophages to lipid-laden macrophages, also known as foam cells (Fig. 1). Hence, the modulation or regulation of the interaction between endothelial and inflammatory cells and the transformation of macrophages to foam cells could be the basis for the beneficial effects of soy-based diets. This review focuses on the atheroprotective effects of soy diets on the four early cellular events contributing to the initiation and progression of atherosclerosis. Since the lipid-lowering effect of soy intake has been discussed previously [25-28], it is not discussed in this review.

1.3. Soy and endothelial cell activation

The cascade of interactions between circulating monocytes and endothelial cells is the prime event in the initiation of atherosclerosis [29,30]. The recruitment of leukocytes, including monocytes, and their adhesion to endothelial cells are a multistep process [29,30] involving initial rolling of monocytes, monocyte activation for subsequent adhesion, and transmigration of monocytes. These steps are controlled by a number of vascular endothelial cell adhesion molecules. Naïve endothelial cells do not promote monocyte adhering to the vascular bed. Initial vascular inflammation results in upregulation of key vascular endothelial cell adhesion molecules such as E- and P-selectins (referred to as CD62E and CD62P, respectively). Inflamed vascular endothelium expressing CD62E and CD62P results in rolling of monocytes on endothelial cells. Rolling is followed by activation of circulating monocytes. Prior activation is a prerequisite for firm adhesion, the next step in the inflammatory processes. Activation of circulating monocytes could be mediated by a number of pro-inflammatory cytokines, oxLDL, and oxLDL/anti-oxLDL IgG immune complexes. Firm adhesion is regulated by vascular cell adhesion molecule-1 (VCAM-1, CD106) and intercellular adhesion molecule-1 (ICAM-1, CD54). The role of cell adhesion molecules in the initiation of inflammatory process has been demonstrated in studies showing that deletion of both CD62E and CD62P has been shown to reduce the progression of atherosclerosis in apoE - / - mice [31 - 33]. Importantly, despite having hypercholesterolemic conditions, double knockout mice lacking the expression of apoE - / - and CD106 have been shown to have reduced numbers of atherosclerotic lesions highlighting the critical role of cell adhesion molecules in the



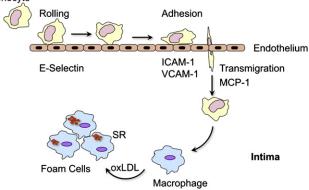


Fig. 1. Inflammatory processes associated with atherosclerosis.

initiation and progression of atherogenesis [34]. Collectively, these studies have demonstrated the causal relationship between adhesion of monocytes to endothelial cells as controlling the initial events of atherogenesis. However, studies on the role of soy in regulation of vascular cell adhesion molecules involved in the inflammatory process associated with atherogenesis are only beginning to emerge.

Pro-inflammatory cytokines. TNF- α and IL-1 β , have been shown to induce activation of endothelial cells resulting in the coordinated up-regulation of CD54, CD62E and CD106 expression [35-37]. Pretreatment of endothelial cells with 17B-estradiol has been shown to inhibit TNF- α -induced up-regulation of vascular cell adhesion molecules [38]. Studies have shown that soy isoflavones may have phytoestrogenic functions [39], suggesting that soy isoflavones may inhibit vascular endothelial cell adhesion molecule expression. Interestingly, endothelial cells treated with genistein, a principal soy isoflavone (25–50 μ M), inhibited TNF- α -induced CD62E and CD106 expression on human umbilical vein endothelial cells and subsequent monocyte adhesion [35,36]. Similar findings were observed using human brain microvascular endothelial cells [40]. An important problem that has not been addressed is whether the *in* vitro findings using pharmacological doses of genistein (50 µM) reflect the in vivo effect of soy-based diets or of soy isoflavones. Recently, Chacko et al. [41,42] have demonstrated genistein at a physiologically relevant concentration (1 µM) inhibited monocyte adhesion to endothelial cells in an in vitro cell culture model. Furthermore, the authors have also provided evidence that peroxisome proliferator-activated receptor (PPAR)-gamma activation contributes to the genistein-mediated inhibition of monocyte adhesion to endothelial cells in cell culture studies [41,42]. Genistein supplementation (2 g/kg diet) has been shown to up-regulate PPAR-alpharegulated genes in vivo [43]. Earlier work from Shay's group [27,44] has shown that modulation of lipid metabolism by soy isoflavone is in part dependent on PPAR-alpha activation in mice fed low (0.09 g/kg diet) vs. high isoflavone (3 g/kg diet) diets. Furthermore, Dang et al. [45] have provided evidence that genistein (at 1 µM) induces PPARgamma activation. Despite these interesting in vitro findings, there is no direct evidence that soy-based diets or soy isoflavones regulate the expression of vascular cell adhesion molecules in vivo. Preliminary findings from our laboratory have shown that expression of CD54, CD62E and CD106 mRNA levels was about 30% to 50% lower in the descending aorta from soy-fed compared with casein-fed mice (unpublished observation). Based on this premise, it is plausible that the atheroprotective effect of soy-based diets in part could be due to inhibition of inflammation-induced CD54, CD62E and CD106 expression (Fig. 2). However, in-depth in vivo or ex vivo studies are warranted to convincingly demonstrate the inhibitory effect of soybased diet on endothelial cell activation associated with chronic inflammatory diseases such as atherosclerosis.

1.4. Soy diet inhibits activation and subsequent adhesion of inflammatory cells

The second step in the inflammatory process is activation and subsequent adhesion of inflammatory cells [23,24]. The adhesive activity of leukocytes is tightly regulated such that circulating monocytes do not adhere to vascular endothelium. However, after activation, the nonadherent monocytes transiently adhere to vascular endothelium. Inflammatory stimuli, such as monocyte chemoattractant protein-1 (MCP-1) and oxLDL, have been shown to activate monocytes, which in turn promote the activated monocytes to firmly adhere to vascular endothelium [46–48]. Moreover, blocking the recruitment of monocytes to sites of inflammation has been shown to reduce lesions and protect animals from atherosclerosis [49,50]. These studies suggest that activation of inflammatory cells is a

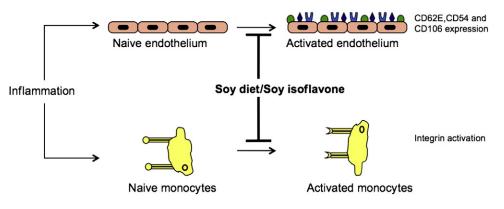


Fig. 2. Soy diet blocks endothelial and monocyte activation.

prerequisite to the progression of the inflammatory processes in atherosclerosis.

Soy feeding has been shown to increase the soy phytochemicals in serum [51,52], suggesting a possibility that the atheroprotective effect of soy isoflavones could also be mediated by blocking inflammatory cell activation and adhesion to vascular endothelium. *In vitro* studies using human monocytic cell lines showed that oxLDL treatment of monocytic cells increases their adhesion to endothelial cells [53]. Furthermore, pre-exposure of monocytes to soy isoflavones (1 µM final concentration with equimolar mixture of genistein, daidzein and equol) as well as to sera from animals fed soy diet inhibited oxLDL-induced monocyte adhesion to endothelial cells [53]. These studies suggest that there is a direct effect of soy isoflavones on monocyte activation and subsequent adhesion, although the detailed mechanism is not known. Based on these findings, it is reasonable to hypothesize that the soy-dependent atheroprotection is mediated through the regulation of monocyte activation (Fig. 2).

1.5. Soy isoflavone regulates monocyte integrin functions

The integrin family of cell surface proteins expressed on inflammatory cells, including monocytes, mediates leukocyte trafficking and adhesion [54,55]. Monocyte integrins on resting cells are not able to bind to their ligands, cell adhesion molecules expressed on vascular endothelium. However, monocyte activation by pro-inflammatory cytokines results in enhanced adhesion of monocytes by regulating the functions of integrins. CD11a, one of the monocyte integrins, is expressed as an inactive low-affinity form [56,57] in circulating (unactivated) monocytes and lymphocytes, and under physiological conditions these cells do not firmly adhere to CD54expressing vascular endothelial cells. Activation of monocytes and T lymphocytes results in the transformation of CD11a from a lowaffinity or avidity to a high-affinity or avidity form [56,57], and such a transformation leads to firm adhesion of inflammatory cells to vascular endothelium through CD11a-CD54 interaction. Recent in vitro studies have shown that pre-exposure of monocytes to soy isoflavones (1 µM) inhibited oxLDL-induced monocyte adhesion to endothelial cells [53]. However, this inhibition of monocyte adhesion is not due to the change in CD11a expression. Instead, soy isoflavones inhibited the transformation of low- to high-affinity form of CD11a, as detected using activation epitope-specific anti-CD11a monoclonal antibody [58,59]. These findings suggest that soy isoflavones inhibit the activation-induced transformation of low- to high-affinity form of the CD11a expressed on monocytes. Inhibition of CD11a-dependent monocyte adhesion to endothelial cells by soy isoflavones could be mediated by two possible mechanisms. Tyrosine kinase inhibitors have been shown to inhibit CD11a binding to CD54 [60]; hence genistein, an isoflavone, with kinase-inhibiting property could have contributed to the inhibition of monocyte adhesion. Furthermore, the phytoestrogenic activity of soy isoflavones [61] may be a contributing factor to the inhibition of monocyte adhesion to CD54. This possibility is supported by a recent report by Friedrich et al. [62] showing that monocytes treated with 17 β -estradiol poorly adhere to endothelial cells. These probable mechanisms warrant further investigation.

Implications of integrin activation in the inflammatory processes are many. For example, monocyte activation and adhesion to vascular endothelium stimulate the expression of tissue factor (TF) on monocytes [63,64]. Specifically, B1-integrin stimulated monocyte adhesion results in up-regulation of monocyte TF expression [65,66]. TF expression has been shown to increase with progression of human vascular disease [67,68] as well as in animal models [69,70]. In addition, B1-integrin-dependent monocyte adhesion leads to induction of pro-inflammatory cytokines, TNF- α and IL-1 β [71,72]. Genistein (20-30 μ M) pretreatment has been shown to inhibit β 1integrin-mediated TF, IL-1 β and TNF- α [65,71,72] expression, suggesting a possible role for soy isoflavones in preventing the integrin-mediated monocyte activation and subsequent adhesiondependent inflammatory responses. Collectively, these studies indicate the significance of integrin-mediated monocyte activation in the inflammatory processes associated with atherosclerosis. These findings further suggest that interfering with leukocyte integrin activation may be a novel concept in the identification of antiinflammatory functions of soy-based diets. However, this concept should be evaluated by determining the expression and activated state of integrins in soy-fed animals using an ex vivo monocyte adhesion model.

1.6. Soy diet modulates transmigration of monocytes

Adhesion of inflammatory cells to vascular endothelium is followed by transendothelial migration of these cells to the arterial intima, the third step in the inflammatory processes. Chemokines that drive leukocyte transendothelial migration have been implicated in the initiation and progression of atherosclerosis. These chemokines include MCP-1, RANTES, fractakline, MIP1- α and MIP1- β . Of these chemokines, MCP-1, a member of the CC chemokines, is characterized by its chemokine activity for inflammatory cells, primarily monocytes [49,50]. MCP-1 is secreted locally by inflamed endothelium and activated monocytes [73,74] and is involved in recruitment of monocytes into arterial walls [74]. Furthermore, studies using CCchemokine receptor 2/apoE double knockout mice have shown that despite having hypercholesterolemic conditions these mice had reduced atherosclerotic lesions [49,50], indicating MCP-1 plays a pivotal role in the inflammatory processes associated with the

pathogenesis and progression of atherosclerosis. Hence, it is reasonable to predict that the atheroprotective effect of soy diet may be mediated by regulating the expression of pro-inflammatory chemokines. Interestingly, soy-based diet inhibits the expression of MCP-1 in the aorta [22], and this observation is also supported by the reduced number of macrophages in atherosclerotic lesion in mice fed sov-based diets compared to control casein diet [22]. Moreover, coculture of human monocytes with endothelial cells resulted in the increased secretion of pro-inflammatory cytokines, such as IL-6 and IL-8, and the addition of soy isoflavones to the monocyte/endothelial cell co-culture inhibited the secretion of these pro-inflammatory cytokines [53]. However, plasma MCP-1 levels were not different in mice fed soy or control (casein) diet [22]. This finding is also supported by recent studies showing there was no change in the plasma inflammation markers, including MCP-1, in postmenopausal women consuming soy isoflavone-enriched foods compared to placebo controls [75]. These findings suggest that soy diets inhibit pro-inflammatory cytokine and chemokine expression at the local inflammation site of lesion formation rather than at the systemic levels of pro-inflammatory cytokines and chemokines. Based on this principle, the atheroprotective effect of soy-based diets may be mediated in part by inhibiting monocyte migration. Such a possibility could be addressed by determining the monocyte migration by bone marrow chimera approach.

1.7. Soy and foam cell formation

After the transmigration of monocytes into arterial intima, monocytes are transformed to macrophages. The activated macrophages further take up oxLDL generated in the intima [76,77] through its binding to scavenger receptors resulting in lipid-laden macrophages, also known as foam cells [78,79]. This step culminates the initial events of atherogenesis. In addition to promoting foam cells, oxLDL binding to endothelial cells leads to up-regulation of cell adhesion molecules such as CD54 and CD106 [80-82]. Furthermore, oxLDL can induce the expression of MCP-1 by vascular endothelial cells, smooth muscle cells and macrophages [83,84], as well as macrophage colony-stimulating factor [85,86]. MCP-1, a monocyte chemokine, initiates monocytes recruitment, and macrophage colony-stimulating factor promotes differentiation of monocytes into macrophages. Owing to the importance of oxLDL generation and scavenger receptor-mediated uptake of oxLDL by macrophages resulting in foam cell formation, the mechanism by which soybased diets can prevent atherosclerosis is in part by inhibiting oxLDL generation. Several in vitro studies have suggested that LDL oxidation is inhibited by soy isoflavones [87-89]. However, components involved in this effect are not consistent between studies. Soy isoflavones, genistein (2.5 µM) and daidzein have been reported to inhibit copper-induced oxidation of LDL [88,89]. However, Kapiotis et al. [87] have reported that only genistein (2.5 µM), not daidzein, inhibits in vitro LDL oxidation. Interestingly, equol and 8-hydroxydaidzein, principal metabolites of daidzein, are potent inhibitors of oxLDL generation than the parent daidzein [89,90]. These in vitro studies have also been supported by in vivo animal studies. Soy feeding or casein diet supplemented with soy isoflavones (7.3 mg isoflavones/kg diet) has been reported to reduce circulating levels of oxLDL and ex vivo LDL oxidation compared with control casein diets in animal models [91,92]. However, clinical studies on the effect of soy isoflavones on oxLDL generation still leave this question unresolved. Following consumption of soy with high isoflavones (210 µM) compared to soy with low isoflavones (7 µM), a decrease in susceptibility of LDL to oxidation has been observed [93]. However, others have reported no effect of soy isoflavones on the generation of oxLDL [94]. The difference could be due to the nature of samples as well as to the nature of assays used to address the antioxidant effects

of soy isoflavones. For instance, the study where the isoflavones have no effect on LDL oxidation is based on samples from hypercholesterolemic patients [94], while the protective effects were reported using samples from individuals with normal serum cholesterol levels [93]. Although there are several reports on soy isoflavone on oxLDL generation, there is no report on the effect of soy isoflavones, particularly genistein, on scavenger receptor expression. Recently, soy pinitol, one of the components of soy, has been shown to reduce foam cell formation in human macrophages by inhibiting the expression of scavenger receptors in a cell culture model [95]. However, more detail studies are warranted to confirm and address possible mechanisms by which soy-based diets regulate macrophage scavenger receptor expression.

1.8. Atheroprotective effect of soy peptides

Soy food or soy protein isolate contains two major components with potential bioactivity. The two components are phytochemicals, such as isoflavones associated with soy protein, and the peptides generated from two of the major soy proteins, such as β -conglycinin (or 7S globulins) and glycinin (or 11S globulins). However, the component(s) of soy responsible for its atheroprotective effects is debatable. Mice fed soy protein isolate that was processed to remove phytochemicals (hereafter referred as isoflavone-free soy protein isolate) had lower incidence of atherosclerotic lesions [22]. Attenuated lesions are also associated with a reduced expression of MCP-1 in aorta and a subsequently lower number of macrophages in lesions in isoflavone-free soy protein isolate-fed than in casein-fed apo $E^{-/-}$ mice [22]. Furthermore, the studies of Adams et al. [21,96] have demonstrated that mice fed isoflavone-free soy protein-containing diet, particularly mice fed the β -conglycinin-containing diet, had a pronounced inhibitory effect on the development of atherosclerosis compared to mice fed casein-lactalbumin-based diets. These findings suggest an atheroprotective role for the protein components of soy diet or peptides generated from soy protein such as β -conglycinin and glycinin. Furthermore, these studies suggest that there are bioactive small peptide fractions produced by the digestion of soy protein that are absorbed from the intestinal tract and have favorable effects on preventing atherosclerosis. One of the caveats in animal experiment testing of the atheroprotective effect of soy diets is the use of caseinfed mice as controls. It is possible that the animal-derived proteins may be atherogenic compared to soy- or other vegetable-derived proteins. However, without a vegetable-based protein diet that may be proatherogenic as controls for the animal studies, the debate on any proatherogenic effect of casein will continue. Alternatively, the atheroprotective effect of soy- or other vegetable-derived proteins reemphasizes the concept that consumption of vegetable-derived proteins may have health-promoting effects not observed with the consumption of animal-derived proteins.

In conclusion, atherosclerosis is a chronic inflammatory disease and the cascades of monocyte-endothelial cell interactions and monocyte migration are crucial processes in the development and progression of atherosclerosis. Adhesion of leukocytes, specifically monocyte adhesion to vascular endothelium, and subsequent transmigration of monocytes to intima resulting in the formation of lipid laden foam cells are initial events in the inflammatory processes associated with atherogenesis. In this review, we have discussed possible mechanism(s) by which soy diet or components of soy diet regulate the inflammatory processes associated with atherosclerosis. Specifically, we have discussed how soy or components of soy diet control adhesive functions of vascular endothelium by regulating key endothelial cell adhesion molecules, monocyte integrin function, cytokines and chemokines that control trafficking and migration of monocytes. Finally, we have discussed a possible regulation of oxLDL generation and regulation of scavenger receptor expression by soy

diets. Thus, nutritional intervention by soy or other diets with antiinflammatory effects could block the inflammatory processes associated with atherogenesis thereby reducing the risk of CVD.

Acknowledgments

We thank John Gregan and Phaedra Yount for their help with manuscript preparation and Dr. John Marecki for the critical review of this article.

References

- D'Armiento FP, Di Gregorio F, Ciafre SA, Posca T, Liguori A, Napoli C, et al. Histological findings and evidence of lipid conjugated dienes and malonyldialdehyde in human fetal aortas. Acta Paediatr 1993;82:823–8.
- [2] Napoli C, D'Armiento FP, Mancini FP, Postiglione A, Witztum JL, Palumbo G, et al. Fatty streak formation occurs in human fetal aortas and is greatly enhanced by maternal hypercholesterolemia. Intimal accumulation of low density lipoprotein and its oxidation precede monocyte recruitment into early atherosclerotic lesions. J Clin Invest 1997;100:2680–90.
- [3] Napoli C, Witztum JL, de Nigris F, Palumbo G, D'Armiento FP, Palinski W. Intracranial arteries of human fetuses are more resistant to hypercholesterolemiainduced fatty streak formation than extracranial arteries. Circulation 1999;99: 2003–10.
- [4] Napoli C, Glass CK, Witztum JL, Deutsch R, D'Armiento FP, Palinski W. Influence of maternal hypercholesterolaemia during pregnancy on progression of early atherosclerotic lesions in childhood: Fate of Early Lesions in Children (FELIC) study. Lancet 1999;354:1234–41.
- [5] Reddy KS. Cardiovascular disease in non-Western countries. N Engl J Med 2004;350:2438–40.
- [6] Lusis AJ, Mar R, Pajukanta P. Genetics of atherosclerosis. Annu Rev Genomics Hum Genet 2004;5:189–218.
- [7] Kolonel LN. Variability in diet and its relation to risk in ethnic and migration groups. Basic Life Sci 1988;43:129–35.
- [8] Adlercreutz H. Western diet and Western diseases: some hormonal and biochemical mechanisms and associations. Scand J Clin Lab Invest 1990;50:3–23.
- [9] Robertson TL, Kato H, Rhoads GG, Kagan A, Marmot M, Syme SL, et al. Epidemiologic studies of coronary heart disease and stroke in Japanese men living in Japan, Hawaii and California. Incidence of myocardial infarction and death from coronary heart disease. Am J Cardiol 1977;39:239–43.
- [10] Hori G, Wang MF, Chan YC, Komatsu T, Wong Y, Chen TH, et al. Soy protein hydrolysate with bound phospholipids reduces serum cholesterol levels inhypercholesterolaemic adult male volunteers. Biosci Biotechnol Biochem 2001;64: 72–8.
- [11] Tonstad S, Smerud K, Hoie LA. comparison of the effects of 2 doses of soy protein or casein on serum lipids, serum lipoproteins, and plasma total homocysteine in hypercholesterolemic subjects. Am J Clin Nutr 2002;76:78–84.
- [12] Jenkins DJ, Kendall CW, Jackson CJ, Connelly PW, Parker T, Faulkner D, et al. Effects of high- and low-isoflavone soyfoods on blood lipids, oxidized LDL, homocysteine, and blood pressure in hyperlipidemic men and women. Am J Clin Nutr 2002;76: 365–72.
- [13] McVeigh BL, Dillingham BL, Lampe JW, Duncan AM. Effect of soy protein varying in isoflavone content on serum lipids in healthy young men. Am J Clin Nutr 2006;83: 244–51.
- [14] Reynolds K, Chin A, Lees KA, Nguyen A, Bujnowski D, He JA. Meta-analysis of the effect of soy protein supplementation on serum lipids. Am J Cardiol 2006;98: 633–40.
- [15] Anthony MS, Clarkson TB, Hughes Jr CL, Morgan TM, Burke GL. Soybean isoflavones improve cardiovascular risk factors without affecting the reproductive system of peripubertal rhesus monkeys. J Nutr 1996;126:43–50.
- [16] Anthony MS, Clarkson TB, Bullock BC, Wagner JD. Soy protein versus soy phytoestrogens in the prevention of diet-induced coronary artery atherosclerosis of male cynomolgus monkeys. Arterioscler Thromb Vasc Biol 1997;17:2524–31.
- [17] Clarkson TB. Nonhuman primate models of atherosclerosis. Lab Anim Sci 1998;48: 569–72.
- [18] Zhang SH, Reddick RL, Piedrahita JA, Maeda N. Spontaneous hypercholesterolemia and arterial lesions in mice lacking apolipoprotein E. Science 1992;258:468–71.
- [19] Plump AS, Smith JD, Hayek T, Aalto-Setala K, Walsh A, Verstuyft JG, et al. Severe hypercholesterolemia and atherosclerosis in apolipoprotein E-deficient mice created by homologous recombination in ES cells. Cell 1992;71:343–53.
- [20] Ni W, Tsuda Y, Sakono M, Imaizumi K. Dietary soy protein isolate, compared with casein, reduces atherosclerotic lesion area in apolipoprotein E-deficient mice. J Nutr 1998;128:1884–9.
- [21] Adams MR, Golden DL, Register TC, Anthony MS, Hodgin JB, Maeda N, et al. The atheroprotective effect of dietary soy isoflavones in apolipoprotein E-/- mice requires the presence of estrogen receptor-alpha. Arterioscler Thromb Vasc Biol 2002;22:1859–64.
- [22] Nagarajan S, Burris RL, Stewart BW, Wilkerson JE, Badger TM. Dietary soy protein isolate ameliorates atherosclerotic lesions in apolipoprotein E-deficient mice potentially by inhibiting monocyte chemoattractant protein-1 expression. J Nutr 2008;138:332–7.

- [23] Ross R. Atherosclerosis-an inflammatory disease. N Engl J Med 1999;340:115-26.
- [24] Libby P. Inflammation in atherosclerosis. Nature 2002;420:868–74.
- [25] Vitolins MZ, Anthony M, Burke GL. Soy protein isoflavones, lipids and arterial disease. Curr Opin Lipidol 2001;12:433–7.
- [26] Clarkson T. Soy, soy phytoestrogens and cardiovascular disease. J Nutr 2002;132: 566S–9S.
- [27] Ricketts ML, Moore DD, Banz WJ, Mezei O, Shay NF. Molecular mechanisms of action of the soy isoflavones includes activation of promiscuous nuclear receptors. A review. J Nutr Biochem 2005;16:321–30.
- [28] Xiao CW, Mei J, Wood CM. Effect of soy proteins and isoflavones on lipid metabolism and involved gene expression. Front Biosci 2008;13:2660–73.
- [29] Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. Cell 1994;76:301–14.
- [30] Galkina E, Ley K. Vascular adhesion molecules in atherosclerosis. Arterioscler Thromb Vasc Biol 2007;27:2292–301.
- [31] Dong ZM, Chapman SM, Brown AA, Frenette PS, Hynes RO, Wagner DD. The combined role of P- and E-selectins in atherosclerosis. J Clin Invest 1998;102: 145–52.
- [32] Ramos CL, Huo Y, Jung U, Ghosh S, Manka DR, Sarembock IJ, et al. Direct demonstration of P-selectin- and VCAM-1-dependent mononuclear cell rolling in early atherosclerotic lesions of apolipoprotein E-deficient mice. Circ Res 1999;84: 1237–44.
- [33] Dong ZM, Brown AA, Wagner DD. Prominent role of P-selectin in the development of advanced atherosclerosis in ApoE-deficient mice. Circulation 2000;101: 2290–5.
- [34] Cybulsky MI, Iiyama K, Li H, Zhu S, Chen M, Iiyama M, et al. A major role for VCAM-1, but not ICAM-1, in early atherosclerosis. J Clin Invest 2001;107:1255–62.
- [35] Weber C, Negrescu E, Erl W, Pietsch A, Frankenberger M, Ziegler-Heitbrock HW, et al. Inhibitors of protein tyrosine kinase suppress TNF-stimulated induction of endothelial cell adhesion molecules. J Immunol 1995;155:445–51.
- [36] May MJ, Wheeler-Jones CP, Pearson JD. Effects of protein tyrosine kinase inhibitors on cytokine-induced adhesion molecule expression by human umbilical vein endothelial cells. Br J Pharmacol 1996;118:1761–71.
- [37] Majewska E, Paleolog E, Baj Z, Kralisz U, Feldmann M, Tchorzewski H. Role of tyrosine kinase enzymes in TNF-alpha and IL-1 induced expression of ICAM-1 and VCAM-1 on human umbilical vein endothelial cells. Scand J Immunol 1997;45: 385–92.
- [38] Caulin-Glaser T, Watson CA, Pardi R, Bender JR. Effects of 17beta-estradiol on cytokine-induced endothelial cell adhesion molecule expression. J Clin Invest 1996;98:36–42.
- [39] Setchell KD. Phytoestrogens: the biochemistry, physiology, and implications for human health of soy isoflavones. Am J Clin Nutr 1998;68:13335–465.
- [40] Lee YW, Lee WH. Protective effects of genistein on proinflammatory pathways in human brain microvascular endothelial cells. J Nutr Biochem 2008;19:819–25.
- [41] Chacko BK, Chandler RT, Mundhekar A, Khoo N, Pruitt HM, Kucik DF, et al. Revealing anti-inflammatory mechanisms of soy isoflavones by flow: modulation of leukocyte-endothelial cell interactions. Am J Physiol Heart Circ Physiol 2005;289:H908–915.
- [42] Chacko BK, Chandler RT, D'Alessandro TL, Mundhekar A, Khoo NK, Botting N, et al. Anti-inflammatory effects of isoflavones are dependent on flow and human endothelial cell PPARgamma. J Nutr 2007;137:351–6.
- [43] Kim S, Shin HJ, Kim SY, Kim JH, Lee YS, Kim DH, et al. Genistein enhances expression of genes involved in fatty acid catabolism through activation of PPARalpha. Mol Cell Endocrinol 2004;220:51–8.
- [44] Mezei O, Li Y, Mullen E, Ross-Viola JS, Shay NF. Dietary isoflavone supplementation modulates lipid metabolism via PPARalpha-dependent and-independent mechanisms. Physiol Genomics 2006;26:8–14.
- [45] Dang ZC, Audinot V, Papapoulos SE, Boutin JA, Lowik CW. Peroxisome proliferatoractivated receptor gamma (PPARgamma) as a molecular target for the soy phytoestrogen genistein. J Biol Chem 2003;278:962–7.
- [46] Jiang Y, Beller DI, Frendl G, Graves DT. Monocyte chemoattractant protein-1 regulates adhesion molecule expression and cytokine production in human monocytes. J Immunol 1992;148:2423–8.
- [47] Carr MW, Alon R, Springer TA. The C-C chemokine MCP-1 differentially modulates the avidity of beta 1 and beta 2 integrins on T lymphocytes. Immunity 1996;4: 179–87.
- [48] Mine S, Tabata T, Wada Y, Fujisaki T, Iida T, Noguchi N, et al. Oxidized low density lipoprotein-induced LFA-1-dependent adhesion and transendothelial migration of monocytes via the protein kinase C pathway. Atherosclerosis 2002;160:281-8.
- [49] Boring L, Gosling J, Cleary M, Charo IF. Decreased lesion formation in CCR2-/mice reveals a role for chemokines in the initiation of atherosclerosis. Nature 1998;394:894-7.
- [50] Gosling J, Slaymaker S, Gu L, Tseng S, Zlot CH, Young SG, et al. MCP-1 deficiency reduces susceptibility to atherosclerosis in mice that overexpress human apolipoprotein B. J Clin Invest 1999;103:773–8.
- [51] Gu L, Laly M, Chang HC, Prior RL, Fang N, Ronis MJ, et al. Isoflavone conjugates are underestimated in tissues using enzymatic hydrolysis. J Agric Food Chem 2005;53:6858–63.
- [52] Gu L, House SE, Prior RL, Fang N, Ronis MJ, Clarkson TB, et al. Metabolic phenotype of isoflavones differ among female rats, pigs, monkeys, and women. J Nutr 2006;136:1215–21.
- [53] Nagarajan S, Stewart BW, Badger TM. Soy isoflavones attenuate human monocyte adhesion to endothelial cell-specific CD54 by inhibiting monocyte CD11a. J Nutr 2006;136:2384–90.

- [54] Hynes RO. Integrins: bidirectional, allosteric signaling machines. Cell 2002;110: 673–87.
- [55] Springer TA, Wang JH. The three-dimensional structure of integrins and their ligands, and conformational regulation of cell adhesion. Adv Protein Chem 2004;68:29–63.
- [56] Tozeren A, Mackie LH, Lawrence MB, Chan PY, Dustin ML, Springer TA. Micromanipulation of adhesion of PMA-stimulated T lymphocytes to planar membranes containing intercellular adhesion molecule-1. Biophys J 1992.
- [57] Kucik DF, Dustin ML, Miller JM, Brown EJ. Adhesion-activating phorbol ester increases the mobility of leukocyte integrin LFA-1 in cultured lymphocytes. J Clin Invest 1996;97:2139–44.
- [58] McDowall A, Leitinger B, Stanley P, Bates PA, Randi AM, Hogg N. The I domain of integrin leukocyte function-associated antigen-1 is involved in a conformational change leading to high affinity binding to ligand intercellular adhesion molecule 1 (ICAM-1). J Biol Chem 1998;273:27396–403.
- [59] Woska Jr JP, Shih D, Taqueti VR, Hogg N, Kelly TA, Kishimoto TK. A small-molecule antagonist of LFA-1 blocks a conformational change important for LFA-1 function. J Leukoc Biol 2001;70:329–34.
- [60] Wang SC, Kanner SB, Ledbetter JA, Gupta S, Kumar G, Nel AE. Evidence for LFA-1/ ICAM-1 dependent stimulation of protein tyrosine phosphorylation in human B lymphoid cell lines during homotypic adhesion. J Leukoc Biol 1995;57:343–51.
- [61] Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, et al. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. Endocrinology 1998;139:4252–63.
- [62] Friedrich EB, Clever YP, Wassmann S, Hess C, Nickenig G. 17Beta-estradiol inhibits monocyte adhesion via down-regulation of Rac1 GTPase. J Mol Cell Cardiol 2006;40:87–95.
- [63] Lo SK, Cheung A, Zheng Q, Silverstein RL. Induction of tissue factor on monocytes by adhesion to endothelial cells. J Immunol 1995;154:4768–77.
- [64] Meisel SR, Xu XP, Edgington TS, Dimayuga P, Kaul S, Lee S, et al. Differentiation of adherent human monocytes into macrophages markedly enhances tissue factor protein expression and procoagulant activity. Atherosclerosis 2002;161:35–43.
- [65] McGilvray ID, Lu Z, Bitar R, Dackiw AP, Davreux CJ, Rotstein OD. VLA-4 integrin cross-linking on human monocytic THP-1 cells induces tissue factor expression by a mechanism involving mitogen-activated protein kinase. J Biol Chem 1997;272: 10287–94.
- [66] Oeth P, Parry GC, Mackman N. Regulation of the tissue factor gene in human monocytic cells. Role of AP-1, NF-kappa B/Rel, and Sp1 proteins in uninduced and lipopolysaccharide-induced expression. Arterioscler Thromb Vasc Biol 1997;17: 365–74.
- [67] Mackman N. Role of tissue factor in hemostasis, thrombosis, and vascular development. Arterioscler Thromb Vasc Biol 2004;24:1015–22.
- [68] Steffel J, Luscher TF, Tanner FC. Tissue factor in cardiovascular diseases: molecular mechanisms and clinical implications. Circulation 2006;113:722–31.
- [69] Hasenstab D, Lea H, Hart CE, Lok S, Clowes AW. Tissue factor overexpression in rat arterial neointima models thrombosis and progression of advanced atherosclerosis. Circulation 2000;101:2651–7.
- [70] Westrick RJ, Bodary PF, Xu Z, Shen YC, Broze GJ, Eitzman DT. Deficiency of tissue factor pathway inhibitor promotes atherosclerosis and thrombosis in mice. Circulation 2001;103:3044–6.
- [71] Lin TH, Rosales C, Mondal K, Bolen JB, Haskill S, Juliano RL. Integrin-mediated tyrosine phosphorylation and cytokine message induction in monocytic cells. A possible signaling role for the Syk tyrosine kinase. J Biol Chem 1995;270: 16189–97.
- [72] Dackiw AP, Nathens AB, Marshall JC, Rotstein OD. Integrin engagement induces monocyte procoagulant activity and tumor necrosis factor production via induction of tyrosine phosphorylation. J Surg Res 1996;64:210–5.
- [73] Weber KS, Draude G, Erl W, de Martin R, Weber C. Monocyte arrest and transmigration on inflamed endothelium in shear flow is inhibited by adenovirusmediated gene transfer of IkappaB-alpha. Blood 1999;93:3685–93.
- [74] Kowala MC, Recce R, Beyer S, Gu C, Valentine M. Characterization of atherosclerosis in LDL receptor knockout mice: macrophage accumulation correlates with rapid and sustained expression of aortic MCP-1/JE. Atherosclerosis 2000;149:323–30.
- [75] Hall WL, Vafeiadou K, Hallund J, Bugel S, Reimann M, Koebnick C, et al. Soyisoflavone-enriched foods and markers of lipid and glucose metabolism in postmenopausal women: interactions with genotype and equol production. Am J Clin Nutr 2006;83:592–600.

- [76] Yla-Herttuala S, Palinski W, Rosenfeld ME, Parthasarathy S, Carew TE, Butler S, et al. Evidence for the presence of oxidatively modified LDL in atherosclerotic lesions of rabbits and man. J Clin Invest 1989;84:1086–95.
- [77] Steinberg D. Low density lipoprotein oxidation and its pathobiological significance. J Biol Chem 1997;272:20963–6.
- [78] Kreiger M. The other side of scavenger receptors: pattern recognition for host defense. Curr Opin Lipidol 1997;8:275–80.
- [79] Greaves DR, Gordon S. Thematic review series: the immune system and atherogenesis. Recent insights into the biology of macrophage scavenger receptors. J Lipid Res 2005;46:11–20.
- [80] Khan BV, Parthasarathy S, Alexander RW, Medford RM. Modified low density lipoprotein and its constituents augment cytokine-activated vascular cell adhesion molecule-1 gene expression in human vascular endothelial cells. J Clin Invest 1995;95:1262–70.
- [81] Takei A, Huang Y, Lopes-Virella MF. Expression of adhesion molecules by human endothelial cells exposed to oxidized low density lipoprotein. Influences of degree of oxidation and location of oxidized LDL. Atherosclerosis 2001;154:79–86.
- [82] Dwivedi A, Anggard EE, Carrier MJ. Oxidized LDL-mediated monocyte adhesion to endothelial cells does not involve NFkappaB. Biochem Biophys Res Commun 2001;284:239–44.
- [83] Lee H, Shi W, Tontonoz P, Wang S, Subbanagounder G, Hedrick CC, et al. Role for peroxisome proliferator-activated receptor alpha in oxidized phospholipidinduced synthesis of monocyte chemotactic protein-1 and interleukin-8 by endothelial cells. Circ Res 2000;87:516–21.
- [84] Miyoshi T, Tian J, Matsumoto AH, Shi W. Differential response of vascular smooth muscle cells to oxidized LDL in mouse strains with different atherosclerosis susceptibility. Atherosclerosis 2006.
- [85] Hamilton JA, Myers D, Jessup W, Cochrane F, Byrne R, Whitty G, et al. Oxidized LDL can induce macrophage survival, DNA synthesis, and enhanced proliferative response to CSF-1 and GM-CSF. Arterioscler Thromb Vasc Biol 1999;19:98–105.
- [86] Sakai M, Biwa T, Matsumura T, Takemura T, Matsuda H, Anami Y, et al. Glucocorticoid inhibits oxidized LDL-induced macrophage growth by suppressing the expression of granulocyte/macrophage colony-stimulating factor. Arterioscler Thromb Vasc Biol 1999;19:1726–33.
- [87] Kapiotis S, Hermann M, Held I, Seelos C, Ehringer H, Gmeiner BM. Genistein, the dietary-derived angiogenesis inhibitor, prevents LDL oxidation and protects endothelial cells from damage by atherogenic LDL. Arterioscler Thromb Vasc Biol 1997;17:2868–74.
- [88] Kerry N, Abbey M. The isoflavone genistein inhibits copper and peroxyl radical mediated low density lipoprotein oxidation in vitro. Atherosclerosis 1998;140: 341–7.
- [89] Turner R, Baron T, Wolffram S, Minihane AM, Cassidy A, Rimbach G, et al. Effect of circulating forms of soy isoflavones on the oxidation of low density lipoprotein. Free Radic Res 2004;38:209–16.
- [90] Kgomotso T, Chiu F, Ng K. Genistein- and daidzein 7-O-beta-D-glucuronic acid retain the ability to inhibit copper-mediated lipid oxidation of low density lipoprotein. Mol Nutr Food Res 2008.
- [91] Damasceno NR, Goto H, Rodrigues FM, Dias CT, Okawabata FS, Abdalla DS, et al. Soy protein isolate reduces the oxidizability of LDL and the generation of oxidized LDL autoantibodies in rabbits with diet-induced atherosclerosis. J Nutr 2000;130: 2641–7.
- [92] Teixeira Damasceno NR, Apolinario E, Dias Flauzino F, Fernandes I, Abdalla DS. Soy isoflavones reduce electronegative low-density lipoprotein (LDL(-)) and anti-LDL (-) autoantibodies in experimental atherosclerosis. Eur J Nutr 2007;46: 125–32.
- [93] Wiseman H, O'Reilly JD, Adlercreutz H, Mallet AI, Bowey EA, Rowland IR, et al. Isoflavone phytoestrogens consumed in soy decrease F(2)-isoprostane concentrations and increase resistance of low-density lipoprotein to oxidation in humans. Am J Clin Nutr 2000;72:395–400.
- [94] Vega-Lopez S, Yeum KJ, Lecker JL, Ausman LM, Johnson EJ, Devaraj S, et al. Plasma antioxidant capacity in response to diets high in soy or animal protein with or without isoflavones. Am J Clin Nutr 2005;81:43–9.
- [95] Choi MS, Lee WH, Kwon EY, Kang MA, Lee MK, Park YB, et al. Effects of soy pinitol on the pro-inflammatory cytokines and scavenger receptors in oxidized lowdensity lipoprotein-treated THP-1 macrophages. J Med Food 2007;10:594–601.
- [96] Adams MR, Golden DL, Franke AA, Potter SM, Smith HS, Anthony MS. Dietary soy beta-conglycinin (7S globulin) inhibits atherosclerosis in mice. J Nutr 2004;134: 511–6.